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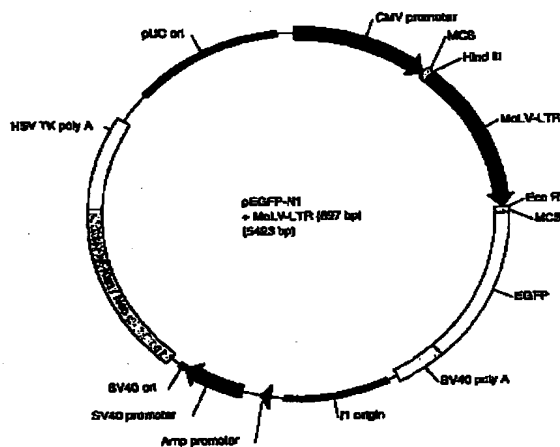
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### (54) Screening method for apoptosis and necrosis

(57) The present invention relates to a method for the determination of non-, anti-, or pro-apoptotic and necrotic conditions of cells, newly designed vectors coding for marker proteins, cell lines transfected with such vector, and a method to assay the non-, pro- or anti-apoptotic or necrotic activity of test compounds.

Figure 3:



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## Description

### Field of the Invention

[0001] The present invention relates to a method for the determination of non-, anti- or pro-apoptotic and necrotic conditions of cells, using newly designed vectors coding for marker proteins, cell lines transfected with such vector, and a method to assay the non-, pro- or anti-apoptotic or necrotic activity of test compounds or physical stimuli.

### Background of the Invention

[0002] Apoptosis plays an essential role in development, i.e. embryogenesis and normal cell turnover, but also in diseases such as cancer, AIDS, neurodegeneration and viral infections. Unlike necrosis, apoptosis is an active, gene-directed self-destruction process of the cell and is associated with characteristic morphological and biochemical changes<sup>1,2</sup>. Nuclear and cytoplasmic condensation and fragmentation of the dying cell into membrane-bound apoptotic bodies are typical characteristics of apoptosis. Another feature of apoptotic cell death is the chromosomal DNA degradation into oligonucleosomal fragments after the activation of specific nucleases<sup>3,4</sup>.

[0003] Apoptosis can be induced by the interaction of the cell surface molecule Fas (CD95) with Fas-Ligand (FasL), where the Fas expressing and sensitive cells undergo apoptosis. Fas is a type I membrane protein, which belongs to the tumor necrosis factor (TNF) and nerve growth factor (NGF) receptor family<sup>5-7</sup>. Fas expression is found on a wide variety of tissues and cells such as thymus, liver, lung, ovary, heart and myeloid cells<sup>9-12</sup>. FasL expression is not only found on lymphocytes but also on a wide variety of tissues and some tumors<sup>13-19</sup>. Both membrane bound FasL and the soluble form (sFasL) can induce apoptosis in Fas positive and sensitive cells. Other forms among a variety of apoptosis mediators include the Perforin/Granzyme system, cytokine deprivation (eg. IL-3 deprivation) and irradiation.

[0004] In contrast to apoptosis, necrosis is a non-physiological death of cells due to chemical or physical injury of the cell membrane. Morphological criteria include cell swelling and cell lysis, lysosomal leakage and loss of the cell membrane integrity.

### State of the Art

[0005] During the last decade, it has become clear that apoptosis plays a keyrole in several diseases. Apoptosis is increased in AIDS, but decreased in cancer and certain autoimmune proliferative diseases.

[0006] Flow cytometry offers a wide variety of possibilities to measure apoptosis. Different methods have been established and implemented, some which stain

on the cell surface and some which stain intracellularly.

[0007] One of the first approach was, beside the observation that apoptotic cells shrink and have higher intracellular granularity, to stain with DNA specific fluorochromes (eg. propidium iodide [PI], ethidium bromide [EtBr]). As soon as a lethal hit is being induced, the DNA starts to change its profile. Apoptotic DNA not only consists of fragmented DNA (visualised as shorter bands, so called DNA ladder, in an agarose gel) but is also partially digested into single nucleotides, so that fluorochromes, like PI or EtBr, have less DNA to stain. This is typically observed by a shift to the left, called sub-G1 peak, on the specific fluorochrome detection chapel in the FACScan™ (from Becton Dickinson, USA). The big disadvantage of this method is that the cell membranes have first to be permeabilized with reagents, like ethanol, in order to stain them with DNA specific dyes, like PI<sup>20</sup>. The treatment is time and labour consuming, and the risks of loosing cells and of handling errors are high. Furthermore, the discrimination between live and apoptotic cells cannot be standardised and requires large experience.

[0008] Another method is the terminal deoxynucleotidyl transferase (TdT)-mediated end-labeling of the DNA strand breaks (TUNEL). The TUNEL method detects DNA strand breaks in cells undergoing apoptosis. TdT is an enzyme which catalyses the addition of deoxyribonucleotide triphosphate to the 3'-OH ends of double or single-stranded DNA. Unlike normal cells, apoptotic cell nuclei incorporate exogenous nucleotides (dUTP)-DIG in the presence of TdT. An anti-DIG antibody fragment with a conjugated fluorochrome enables the visualisation of apoptotic cells. An increase of apoptotic cells causes a higher number of DNA fragments and consequently a brighter fluorescence. An advantage of this method is the very high specificity<sup>21</sup>. A disadvantage of this method is that it is expensive and can only be used for a small set of samples, because it is time intensive. Therefore, it is not applicable for large screening programmes.

[0009] The loss of cell membrane polarity and the presentation of increased amounts of phosphatidyl serine (PS) on the outside of the cell membrane during the early phase of apoptosis has led to yet a new approach. Annexin V is a calcium-dependent phospholipid-binding protein with high affinity for PS. Due to the fact, that the cell membrane integrity is maintained in the early and intermediate phases of apoptosis but not in necrosis, it is possible to distinguish between apoptotic and necrotic cells, when Annexin V is used concomitantly with the DNA dye PI. Early and intermediate apoptotic cells show increased binding of Annexin-FITC and are mainly negative for PI-staining. Late apoptotic stages and necrotic cells become double positive, because of PS presentation on the surface and the PI staining of intracellular nucleic acids due to disintegration of the membrane<sup>22</sup>. This method is also costly and labour intensive.

[0010] Green fluorescence protein (GFP) from the jellyfish *Aequorea victoria* can be used to monitor gene expression and protein localisation in living organisms (in vivo) and in vitro<sup>23-26</sup>. GFP-fluorescence is stable, can be monitored noninvasively in living cells and persists in paraformaldehyde-fixed cells. FACS-optimized mutants of green fluorescence protein have been developed<sup>8</sup>. One of these mutants (GFPmut1) has been integrated into the pEGFP vectors and is commercially available (from Clontech). The big advantage of this mutant is that the maximal excitation peak of GFPmut1 is 488 nm and the emission maxima is 507 nm. Conventional flow cytometers are equipped with an argon laser emitting light at 488 nm and have the suitable detection filters already installed, making the GFPmut1-protein an ideal candidate for flow cytometry studies and fluorescence microscopy.

[0011] GFP has been used as a marker protein to detect cells transiently transfected with the commercially available plasmid pEGFP-C1 (Clontech)<sup>24</sup>. According to this publication apoptosis was detected by reduced fluorescence of the DNA-binding dye PI in the apoptotic subpopulation. It was not recognized that GFP itself could be used as a marker for apoptosis.

[0012] Experiments by the present inventors have shown, that stable transfection of eukaryotic cells (e.g. A20.2J) with the pEGFP-C1 (as received from the manufacturer Clontech with the cytomegalovirus CMV promoter) results in little or no expression of the GFPmut1 gene.

[0013] For the foregoing reason there is a need for a new and improved method and tools for determining apoptosis and necrosis of cell lines which in particular can be used to efficiently and cheaply assay compounds on their pro- or anti-apoptotic activity.

#### Objective of the invention

[0014] Accordingly, it is the objective of the invention to provide an improved method for the determination of apoptotic and/or necrotic conditions of cells and to provide improved tools useful for this method.

#### Summary

[0015] Our invention is directed to a method for the determination of apoptotic and/or necrotic conditions of living test cells that satisfies the hereinbefore discussed needs. This method comprises monitoring the presence of a marker protein in said cells, in particular, wherein the apoptotic and/or necrotic condition is monitored in the presence of a non-, pro- or anti-apoptotically or necrotically active compound, and/or a physical stimulus, and wherein the marker protein is preferentially produced in the test cells after stable transfection of said cells with a DNA coding for and expressing the marker protein.

[0016] A preferred marker protein is the Green Fluorescent Protein (GFP) or a fluorescent mutant thereof, e.g. GFPmut1.

[0017] Another embodiment of our invention is directed to a new vector for the stably transfection of test cells which are able to highly express said marker protein. We found that the known hEF1- $\alpha$  promoter, and the new combination of the CMV and the MoLV-LTR promoter are particularly well suited for enhancing transcription of the GFPmut1 gene.

[0018] Another embodiment of our invention is directed to a live cell line stably transfected with such vector. Such cell line can be used in an assay to determine non-, pro- or anti-apoptotic or necrotic activity of test compounds.

[0019] Another embodiment of our invention is directed to a method to assay the non-, pro- or anti-apoptotic or necrotic activity of a test compound on live test cells. This method comprises stably transfecting a group of said cells with a vector coding for and expressing a marker protein. The transfected cells are treated in a suitable culture medium with the test compound. The presence, i.e. the decrease or increase, respectively, of the expressed marker protein in said group of cells is monitored by conventional methods, and compared with the results observed with a parallel group of the same test cells which was not exposed to the test compound. The test compound may consist of a multiplicity of compounds, e. g. as obtained from combinatorial chemistry methods. Of particular interest are the apoptotic or necrotic conditions of normal and cancer cells under the influence of test compounds and/or physical stimuli.

[0020] These and other features, aspects, and advantages of our invention will become better understood with reference to the following description, appended claims and accompanying drawings.

#### Short Description of the Drawings

[0021]

Fig. 1: pBluescriptIIKS(+)-EF-1 $\alpha$  plasmid. The promoter of human EF-1 $\alpha$  has been cloned into the pBluescriptIIKS(+)-vector (from Stratagene) with Hind III and Xba I.

Fig. 2: pBluescriptIIKS(+)-EF-1 $\alpha$ -GFP plasmid. The gene coding for GFPmut1 (from pEGFP-C1, Clontech) has been cloned into the pBluescriptIIKS(+)-EF-1 $\alpha$  plasmid with Xba I and Ssp I.

Fig. 3: pEGFP-N1+MoLV-LTR plasmid. The MoLV-LTR promoter has been cloned into the pEGFP-N1 vector (from Clontech) with Hind III and Eco RI.

Fig. 4: FACS-histogramm showing the difference of the fluorescing capacities between non-transfected living A20 and GFP-transfected living A20GFP measured on the FL-1 channel.

Fig. 5: FACS-histogramm showing the difference of the fluorescing capacities between non-transfected living PB3c and GFP-transfected living PB3cGFP

measured on the FL-1 channel.

Fig. 6: FACS-histogramm showing the difference of the fluorescing capacities between non-transfected living Jurkat and GFP-transfected living JurkatGFP measured on the FL-1 channel.

Fig. 7a: FACS dot plot showing the cell size (FSC) and granularity (SSC) of living A20GFP.

Fig. 7b: FACS-histogramm showing the typical GFP-associated fluorescence profile of living A20GFP.

Fig. 8a: FACS dot plot showing the shrinkage and increased granular density (compared to Fig. 7a) of sFasL SN treated and therefore mainly apoptotic A20GFP.

Fig. 8b: FACS-histogramm showing the decrease of GFP associated fluorescence in sFasL SN treated and therefore mainly apoptotic A20GFP.

Fig. 8c: FACS dot plot showing the difference between high fluorescing living A20GFP (R1) and low fluorescing apoptotic A20GFP (R2).

Fig. 8d: FACS dot plot of high fluorescing living A20GFP (gate R1 from Fig. 8c). The high fluorescing A20GFP cells (R1) belong mainly to the not shrunken and not granular dense cell population (typical pattern of living cells).

Fig. 8e: FACS dot plot of low fluorescing apoptotic A20GFP (gate R2 from fig. 8c). The low fluorescing A20GFP cells belong mainly to the shrunken and granular dense cell population (typical feature of apoptotic cells).

Fig. 9a: FACS dot plot showing the cell size and granularity pattern of living A20GFP (ethanol treated and PI-stained).

Fig. 9b: FACS-histogramm showing the typical DNA-pattern of PI-stained living A20GFP.

Fig. 10a: FACS dot plot showing the shrinkage of sFasL SN treated A20GFP (ethanol treated and PI-stained).

Fig. 10b: FACS-histogramm showing the fragmentation of DNA (represented by the sub-G1 peak = M1), as proof for apoptosis in sFasL SN treated A20GFP.

Fig. 11: FACS-histogramm showing the fluorescence intensity of living A20GFP compared to apoptotic A20GFP, necrotic A20GFP and living non-transfected A20.

Fig. 12a: FACS-histogramm showing the induction of apoptosis by sFasL SN in A20GFP (control experiment to Fig. 12b).

Fig. 12b: FACS-histogramm showing the induction of apoptosis by sFasL SN in A20GFP in the presence of ActD and CHX. Neither a reduction of GFP-associated fluorescence nor a inhibition of apoptosis can be observed.

Fig. 13: FACS-histogramm showing the anti-apoptotic effect of IL-3 treatment. Apoptotic PB3cGFP cells show a lower GFP-associated fluorescence intensity compared to the IL-3 treated and therefore

living PB3cGFP.

Fig. 14: FACS-histogramm showing the pro-apoptotic effect of sFasL SN on JurkatGFP, visualised as a decrease of GFP-associated fluorescence in apoptotic JurkatGFP cells compared to living JurkatGFP

#### Description

[0022] Apoptotic and/or necrotic conditions of cells can be induced either physically, e.g. by irradiation, or chemically, e.g. by treatment with pro-apoptotically or necrotically or other active compounds.

[0023] Marker proteins, that can be used for the method, are proteins which are detected indirectly by staining or directly by their physical and/or chemical characteristics, e.g. by fluorescence. They can be directly introduced into the cells or produced internally after transfection with a vector expressing the marker protein. Direct introduction of the marker protein may be achieved by methods known in the art, e.g. by the use of liposomes. Test cells are transfected either transiently, e. g. with a lipid such as the LipofectAMINE™ reagent, or preferentially stably by use of a gene-gun or other suitable methods, and preferably by electroporation. Useful vectors are of RNA or preferably of DNA origin. The marker protein is produced in the test cells after transfection with the vector, usually a DNA vector, coding for and expressing the marker protein. Useful marker proteins are known and comprise any detectable proteins, in particular fluorescent proteins, e.g. the blue fluorescent protein (BFP), a green fluorescent protein such as GFPuv or GFPwt. A preferred marker protein is the GFPmut 1.

[0024] The amount of intact marker protein (marker function) present in the cells can either be visualized indirectly by staining, e.g. with antibodies, or directly by the physical and/or chemical characteristics of the marker protein itself, preferably by fluorescence. Under necrotic conditions the marker protein loses its function much faster and more completely compared to apoptotic conditions. Under necrotic condition the marker function is lost within a few hours, whereas functional loss is delayed under apoptotic conditions. The difference in time also allows the distinction between these two cell conditions.

[0025] Useful visualisation is performed on fixed, e.g. in paraformaldehyde, or live cells, but monitoring is preferably performed on live cells in the culture medium either by color reactions or preferentially by fluorescence measurements. Detection devices comprise flow cytometry, e.g. FACScan™, fluorescence platereaders, e.g. FLIPR™, or any other suitable detection device. The acquisition device used is a FACScan™. The following parameters are preferentially measured:

- FSC-Height (forward scatter) as a measure for the cell size,

- SSC-Height (side scatter) as a measure for the internal granularity (density) of the cells,
- green fluorescence (e.g. of GFPmut1) is visualised on the FL-1 channel (FL-1 Height) in the FACS-can™.

[0026] The results are compared after dot plot and/or histogram visualisation (see Fig. 4 to 14).

[0027] Useful test cells comprise eukaryotic and prokaryotic cells. The test cells which are used in the method are cells of any type, preferentially normal, infected, e. g. with virus, parasites, bacteria or prions infected, tumor cells or genetically manipulated cells of human or animal origin, which can be cultured *in vitro*, carrying a vector expressing the marker gene. Such test cells are for example lymphoma cells, e.g. A20.2J, Jurkat, or mast cells, e.g. PB-3c, which are transfected with DNA vectors pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP or pEGFP-N1+MoLV-LTR, respectively, to give the live cell lines named A20GFP, PB3cGFP or JurkatGFP, respectively.

[0028] In a preferred method the presence of Green Fluorescent Protein (GFP) or a fluorescent mutant thereof (e.g. GFPmut1) is monitored by means of a flow cytometer measuring the fluorescence intensity. In general two groups of test cells are used, each of a defined number of cells (e.g. cancer cells), which are transfected with a DNA vector according to our invention. Depending on the detection device the number of test cells in the test tubes is 5 to 50'000'000, but for flow cytometry 10'000 to 500'000 are preferably used.

[0029] In a preferred method two groups of test cells are used, each of a defined number of cells (e.g. cancer cells), which are stably transfected with any of the vectors according to the invention, incubating one group together with the test compound in a culture medium, stimulating the cells of both groups with an excitation beam, usually with a wave length of high energy, in particular an argon laser, determining the fluorescing intensity of the cells of each group by means of a flow cytometer, presently on the FL-1 channel (FL-1 Height), and comparing the obtained fluorescing intensity of the cells of the two groups.

[0030] Another embodiment of the invention is directed to vectors useful in the method described above. A vector according to the invention comprises a gene coding for the marker protein which is operably linked to one or more strong promoters, preferably the hEF1- $\alpha$  promoter, the MoLV-LTR promoter or a combination of the CMV and the MoLV-LTR promoter. The promoter of EF-1 $\alpha$  (elongation factor-1 $\alpha$ ) chromosomal gene efficiently stimulates *in vitro* transcription<sup>28</sup>. EF-1 $\alpha$  promotes the GTP-dependent binding of an aminoacyl-tRNA to ribosomes in eukaryotic cells. EF-1 $\alpha$  is one of the most abundant proteins in eukaryotic cells and is expressed in almost all kinds of mammalian cells. The MoLV-LTR (Moloney Leukemia Virus-Long Terminal Repeat) is a very effective promoter<sup>33</sup>. These findings

make the promoter of EF-1 $\alpha$  and the MoLV-LTR promoter preferred candidates for enhancing transcription of a desired gene.

[0031] Preferred vectors have a gene coding for the GFP or a fluorescent mutant thereof. They are in particular able to express the marker protein GFPmut1. Such vectors are specifically the plasmid pBluescript II KS(+)-EF-1 $\alpha$ -EGFP and pEGFP-N1+MoLV-LTR.

[0032] Advantageously vectors with one or more strong promoters are used in order to obtain a high expression of the gene coding for the marker protein, preferably GFPmut1.

[0033] The present vectors are produced by methods known in the art.

[0034] Another embodiment of the invention is directed to live cell lines useful in the above method. Preferred living cell lines are transfected with a preferred vector according to the invention and are named A20GFP, PB3cGFP or JurkatGFP.

[0035] A20.2J is preferentially transfected with the pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP vector, whereas Jurkat and PB-3c are transfected with the pEGFP-N1+MoLV-LTR vector. The cells are preferably transfected by electroporation. The transfected cells are selected by single cell assay in order to obtain clonally and stably transfected cell lines. The selection process involves high GFP expression (high fluorescence intensity) and GFP-fluorescence of the selected cells in a narrow range, in order to obtain a homogenous high GFP-expressing cell line.

[0036] In another embodiment the invention is directed to an assay to determine the non-, pro-, or anti-apoptotic or necrotic activity of new or known test compounds. The test is performed in that a defined number of transfected and selected cells is incubated with a test compound or/and a physical stimuli, e.g. sonication or irradiation, in the appropriate culture medium. Preferentially 250'000 cells per condition are used. Any culture medium can be used in which the cells survive and/or grow. Preferred culture media comprise e.g. Iscove's for A20GFP, Iscove's plus IL-3 for PB3cGFP, and RPMI for JurkatGFP.

[0037] For comparative and standardisation purposes useful apoptotic compounds are e. g. proteins, such as FasL, fatty acids, e.g. ceramide, steroids, e.g. cortisol, antibiotics, e.g. bleomycin, and others. A preferred apoptotic substance for A20GFP and JurkatGFP is FasL. A useful anti-apoptotic substance for PB3cGFP is IL-3. Toxicity and thereby necrosis can be induced with a wide variety of substances. Preferred induction of necrosis includes the use of antibodies plus complement on A20GFP.

[0038] Cells can be cultured at different temperatures, but for the assay the test cells are preferably cultured at 37 °C during the time of the assay. In FACS-analysis 10 000 cells are usually evaluated from a pool of 250 000 cells.

[0039] Live, transfected cells, in comparison to apop-

totic transfected cells, show a high level of specific fluorescence intensity, whereas apoptotic cells lose part of their fluorescent capacity after some hours (shift to the left on the fluorescence detecting channel FL-1). On the other hand, necrotic cells lose all their fluorescing capacity within a couple of hours and at that time render them indistinguishable from non-transfected cells.

[0040] The reduction of the experimental set-up to a single parameter (a marker protein, e. g. GFPmut1) and the possibility of standardisation renders the present invention a useful and improved method for the screening of non-, pro-, antiapoptotic and necrotic substances and/or physical stimuli. Standardisation is possible for the transfection and selection method, the fluorescence measuring, the numbers of transfected cells, the amount of test compounds used, the type and intensity of the physical stimuli, and other standardizable parameters, such as the setup of the flow cytometer and/or any other suitable screening device, e. g. fluorescence platereaders.

[0041] Although the present invention has been described in considerable detail with reference to certain preferred embodiments thereof, it should be noted that other embodiments or versions are possible. For example for the detection of non-, pro- or anti-apoptotically or necrotically active compounds on specific cell lines, e. g. specific cancer cell lines, such as cell lines of malignant tumors of human breast, lung, brain, colon, prostate and the like, the relevant cell line may be stably transfected with a vector of the invention, or vectors encoding other easily detectable proteins may be constructed and used for transfection. Therefore the spirit and scope of the appended claims should not be limited to the description of the preferred versions contained therein.

[0042] The following examples describe the invention in still more detail but they also should not be construed as a limitation of the present invention.

#### Example 1: Construction of pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP

[0043] For the construction of the pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP plasmid (see Fig. 2) the promoter of human EF-1 $\alpha$  (from pEF-BOS, kindly provided by S. Nagata<sup>27</sup>) and the gene EGFP + SV40 poly A coding for GFPmut1 (from pEGFP-C1, Clontech) were cloned into the pBluescriptIIKS(+)-vector (from Stratagene).

[0044] The first step was to clone the promoter of human EF-1 $\alpha$  (from pEF-BOS) into the Xba I and Hind III predigested pBluescriptIIKS(+) vector.

[0045] The resulting EF-1 $\alpha$  promoter fragment, after Hind III and Xba I digestion of pEF-BOS, has a length of 1188 bp, whereas the other resulting fragment corresponds to the pEF-BOS backbone.

[0046] The pBluescriptIIKS(+) contains a unique Xba I restriction site (position 677) and a unique Hind III restriction site (position 719). The two resulting frag-

ments after Xba I and Hind III digestion have a length of 2919 bp and 42 bp, respectively.

[0047] Three  $\mu$ g pEF-BOS vector were mixed in an Eppendorf tube (tube a) with 1  $\mu$ l Hind III (from Boehringer Mannheim, 12 U/ $\mu$ l), 1  $\mu$ l Xba I (from New England Biolabs, 20000 U/ml), 3  $\mu$ l 10x incubation buffer for restriction enzyme B (from Boehringer Mannheim) and 20  $\mu$ l double distilled water making a final volume of 30  $\mu$ l. In another Eppendorf tube (tube b), 3  $\mu$ g pBluescriptIIKS(+) were pipetted together with 1  $\mu$ l Hind III, 1  $\mu$ l Xba I, 3  $\mu$ l 10x incubation buffer for restriction enzyme B and 20  $\mu$ l double distilled water to make a final volume of 30  $\mu$ l. Both Eppendorf tubes (tubes a and b) were incubated for 3 hours at 37°C. After incubation, 11  $\mu$ l DNA-loading buffer (0.25% bromophenol blue from Sigma, 0.25% xylene cyanol from Sigma and 15% Ficoll type 400 from Pharmacia in water) were added to both tubes and thereafter the contents were loaded onto separate slots on a 0.6% low-melting-point (LMP) agarose (from Bethesda Research Laboratories) gel. In a third slot, 9  $\mu$ l 1 Kb DNA-ladder (from Promega), diluted 1:3 with DNA-loading buffer, were loaded on the gel. The 0.6% LMP agarose gel is prepared by mixing 0.3 g LMP-agarose were dissolved in 50 ml 1x TAE (50x TAE concentrated stock solution /per liter): 242 g Tris(hydroxymethyl)-aminomethane (from Merck), 57.1 ml glacial acetic acid (from Merck), 100 ml 0.5 M EDTA (from Sigma) (pH 8.0) and double distilled water were added to a final volume of 1 liter), heating the mixture until it is dissolved, and adding ethidiumbromide solution (from Sigma) to a final concentration of 1  $\mu$ g/ml. The gel is run at 60-70 Volts for 60-90 minutes. The ethidiumbromide stained DNA-bands in the gel are visualized with 300 nm UV-light and their size are determined relative to the 1 Kb DNA-ladder. The DNA-band containing the 1188 bp long fragment (EF-1 $\alpha$  promoter) from the digested pEF-BOS vector and the DNA-band containing the 2919 bp long fragment from the digested pBluescriptIIKS(+) were cut out from the LMP agarose gel and put into two different Eppendorf tubes. Both tubes were centrifuged briefly and incubated at 65 °C for 5 minutes. Afterwards, 3.5  $\mu$ l of the 2919 bp long fragment (Hind III and Xba I digested pBluescript II KS(+)) were mixed together with 7  $\mu$ l of the 1188 bp long fragment (EF-1 $\alpha$  promoter from the Hind III and Xba I digested pEF-BOS) in a separate Eppendorf tube, incubated at 65 °C for 5 minutes and centrifuged briefly. The mixture was cooled at 37 °C and then the following ligation mix was added: 2  $\mu$ l 10x ligation buffer (from Appligene, provided with T4 DNA ligase), 7.5  $\mu$ l double distilled water and 1  $\mu$ l T4 DNA ligase (from Appligene, 5 U/ $\mu$ l). The ligation reaction was incubated overnight at 15 °C.

[0048] The newly ligated pBluescriptIIKS(+)-EF-1 $\alpha$  was transformed into One Shot™ competent *E. coli* (from Invitrogen), according to the manufacturer protocol of Invitrogen.

[0049] Ten  $\mu$ l of the ligation reaction were used for the

transformation of competent *E. coli*. 50, 100 and 200  $\mu$ l of the transformation mixture were plated on different ampicillin-containing LB agar plates (10 g tryptone, 5 g yeast extract, 10 g NaCl, 20 g bactoagar (from Difco Laboratories Detroit Michigan USA), adjust pH to 7.0 with NaOH, water to a final volume of 1 liter, autoclave, cool the agar-media to 55 °C, add ampicillin solution to a final concentration of 50  $\mu$ g/ml and pour the plates with 10-15 ml agar/plate). The plates/transformation mix were then incubated at 37 °C overnight. The next day different bacterial colonies were picked for further analysis of recombinant DNA clones by small-scale plasmid DNA isolation (according to the alkaline lysis method described in Maniatis *et al.* <sup>32</sup>).

[0050] To check the presence of the EF-1 $\alpha$  promoter fragment cloned into pBluescript II KS(+), a Hind III and Xba I restriction enzyme digestion of the isolated DNA was carried out (3  $\mu$ l 10x incubation buffer for restriction enzyme B, 5  $\mu$ l plasmid DNA from the alkaline lysis method, 22  $\mu$ l double distilled water, 1  $\mu$ l Hind III, 1  $\mu$ l Xba I were mixed together in an Eppendorf tube and incubated for 3 hours at 37 °C), loaded on a 1% agarose (from Bio-Rad) gel and subsequently visualized with 300 nm UV-light and analyzed. The expected fragment of 1188 bp length after Hind III and Xba I digestion was observed, and therefore confirmed the presence of the cloned fragment (see also Fig. 1).

[0051] In order to obtain large quantities of the newly created plasmid, a large-scale plasmid DNA isolation was performed according to the Qiagen Plasmid Maxi Protocol.

[0052] The second step was to clone the EGFP + SV40 poly A (from pEGFP-C1) into the Xba I and Ssp I predigested pBluescriptIIKS(+)-EF-1 $\alpha$ .

[0053] The pEGFP-C1 vector (from Clontech) contains a unique Nhe I restriction site (position 592) and two Ssp I restriction sites (position 1664 and 2217). The three resulting fragments, after Nhe I and Ssp I digestion, have a length of 3106 bp, 1072 bp and 553 bp, respectively.

[0054] The pBluescriptIIKS(+)-EF-1 $\alpha$  (see Fig. 1) contains an Xba I and two Ssp I restriction sites and the three resulting fragments, after Xba I and Ssp I digestion, have a length of 3319 bp, 658 bp and 130 bp, respectively.

[0055] Of the pEGFP-C1 vector 4.2  $\mu$ g were mixed in an Eppendorf tube (tube 1) with 1  $\mu$ l Nhe I (from New England Biolabs, 4000 U/ml), 1  $\mu$ l Ssp I (from Boehringer Mannheim, 10 U/ $\mu$ l), 3  $\mu$ l 10x incubation buffer for restriction enzyme M (from Boehringer Mannheim) and 18  $\mu$ l double distilled water making a final volume of 30  $\mu$ l. In another Eppendorf tube (tube 2), 3-4  $\mu$ g pBluescriptIIKS(+)-EF-1 $\alpha$  were pipetted together with 1  $\mu$ l Xba I, 1  $\mu$ l Ssp I, 3  $\mu$ l 10x incubation buffer for restriction enzyme H (from Boehringer Mannheim) and 18  $\mu$ l double distilled water to make a final volume of 30  $\mu$ l. Both Eppendorf tubes (tubes 1 and 2) were incubated for 4 hours at 37°C. After incubation, 11  $\mu$ l DNA-loading

buffer was added to both tubes and thereafter the contents were loaded into separate slots on a 0.6% low-melting-point (LMP) agarose gel. In a third slot, 9  $\mu$ l 1 Kb DNA-ladder, diluted 1:3 with DNA-loading buffer, were loaded on the gel. The gel was run at 60-70 Volts for 60-90 minutes. The ethidiumbromide stained DNA-bands in the gel were visualized with 300 nm UV-light and their size was determined relative to the 1 Kb DNA-ladder. The DNA-band containing the 1072 bp long fragment from the digested pEGFP-C1 vector and the DNA-band containing the 3319 bp long fragment from the digested pBluescriptIIKS(+)-EF-1 $\alpha$  were cut out from the LMP agarose gel and put into two different Eppendorf tubes. Both tubes were briefly centrifuged and incubated at 65 °C for 5 minutes. Afterwards, 3.5  $\mu$ l of the 3319 bp long fragment (Xba I and Ssp I digested pBluescript II KS(+) + EF-1 $\alpha$ ) were mixed together with 7  $\mu$ l of the 1072 bp long fragment (EGFP + SV40 poly A from the Nhe I and Ssp I digested pEGFP-C1) in a separate Eppendorf tube, incubated at 65 °C for 5 minutes and centrifuged briefly. The mixture was cooled at 37 °C and then the following ligation mix was added: 2  $\mu$ l 10x ligation buffer, 7.5  $\mu$ l double distilled water and 1  $\mu$ l T4 DNA ligase. The ligation reaction was incubated overnight at 15 °C.

[0056] The newly ligated pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP was transformed into One Shot™ competent *E. coli* (from Invitrogen), according to the manufacturer protocol of Invitrogen. Ten  $\mu$ l of the ligation reaction were used for the transformation of competent *E. coli*. 50, 100 and 200  $\mu$ l of the transformation mixture were plated on different ampicillin - containing LB agar plates and incubated at 37 °C overnight. The next day different bacterial colonies were picked for further analysis of recombinant DNA clones by small-scale plasmid DNA isolation (according to the alkaline lysis method described in Maniatis *et al.* <sup>7</sup>).

[0057] The newly constructed pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP vector (see Fig. 2) contains two Sal I restriction sites, one in the pBluescriptIIKS(+) backbone and the other one in the cloned EGFP+SV40 poly A fragment. The two resulting fragments after a Sal I digestion have a length of 1981 bp and 2410 bp, respectively.

[0058] In order to check the presence of the EGFP + SV40 poly A fragment cloned into pBluescriptIIKS(+)-EF-1 $\alpha$ , a Sal I restriction enzyme digestion of the isolated DNA was carried out (3  $\mu$ l 10x incubation buffer for restriction enzyme H, 5  $\mu$ l plasmid DNA from the alkaline lysis method, 22  $\mu$ l double distilled water, 1  $\mu$ l Sal I (from Appligene, 10 U/ $\mu$ l) were mixed together in an Eppendorf tube and incubated for 3 hours at 37 °C), loaded on a 1% agarose gel and subsequently visualized with 300 nm UV-light and analyzed. The two expected fragments of 1981 bp and 2410 bp length after Sal I digestion were observed, and therefore confirmed the presence of the cloned fragment.

[0059] In order to obtain large quantities of the newly created plasmid (see figure 2), a large-scale plasmid

DNA isolation has been performed according to the Qia-gen Plasmid Maxi Protocol.

[0060] At this point, to check the functionality of the newly constructed pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP plasmid, COS-1 cells (ATCC CCL 1850) (COS-1 is a fibroblast-like cell line established from CV-1 simian cells from ATCC CRL 1850) were transiently transfected with 2  $\mu$ g of the plasmid, according to the DEAE-Dextran Transfection Protocol from the Mammalian Transfection Kit of Stratagene. After 1-3 days incubation at 37 °C, the transiently transfected COS-1 cells were analyzed under the fluorescence microscope and highly fluorescing GFP-positive COS-1 cells have been observed, indicating that the constructed GFP-carrying plasmid was functional.

#### Example 2: Construction of pEGFP-N1+MoLV-LTR

[0061] For the construction of the pEGFP-N1+MoLV-LTR plasmid (see Fig. 3) the MoLV-LTR promoter<sup>33</sup> (from Gem-MoLV-LTR) was cloned into the Hind III and Eco RI predigested pEGFP-N1 vector (from Clontech).

[0062] The MoLV-LTR promoter<sup>33</sup> was cloned with Hind III and Eco RI into the pGEM-3Zf(+) vector (from Promega). The Gem-MoLV-LTR contains a unique Hind III and a unique Eco RI restriction site. The resulting MoLV-LTR promoter fragment, after Eco RI and Hind III digestion, has a length of 697 bp, whereas the other resulting fragment corresponds to the pGEM-3Zf(+) backbone.

[0063] The pEGFP-N1 (from Clontech) contains a unique Hind III restriction site (position 623) and a unique Eco RI restriction site (position 630). The two resulting fragments, after Hind III and Eco RI digestion, have a length of 4726 bp and 7 bp, respectively.

[0064] Of pEGFP-N1 7.3  $\mu$ g were pipetted into an Eppendorf tube (tube A) together with 1  $\mu$ l Hind III, 3  $\mu$ l 10x incubation buffer for restriction enzyme B (from Boehringer Mannheim) and 21  $\mu$ l double distilled water to make a final volume of 30  $\mu$ l. The tube was incubated for 3 hours at 37 °C. Afterwards, it was incubated at 65 °C for 5 minutes, cooled down at 37 °C and 9  $\mu$ l double distilled water, 1  $\mu$ l 10x incubation buffer for restriction enzyme B and 1  $\mu$ l Eco RI were added to the mixture making a final volume of 40  $\mu$ l. In another Eppendorf tube (tube B), 3  $\mu$ g Gem-MoLV-LTR were mixed with 1  $\mu$ l Hind III, 1  $\mu$ l Eco RI (from Boehringer Mannheim, 10 U/ $\mu$ l), 3  $\mu$ l 10x incubation buffer for restriction enzyme B and 20  $\mu$ l double distilled water making a final volume of 30  $\mu$ l. Both Eppendorf tubes (tubes A and B) were incubated for 3 hours at 37 °C. After incubation, 14  $\mu$ l DNA-loading buffer were added to both tubes and thereafter the contents were loaded into separate slots on a 0.6% low-melting-point (LMP) agarose gel. In a third slot, 15  $\mu$ l 1 Kb DNA-ladder (from Promega), diluted 1:3 with DNA-loading buffer, were loaded on the gel. The gel was run at 60-70 Volts for 60-90 minutes. The ethidium-bromide stained DNA-bands in the gel were visualized

with 300 nm UV-light and their size were determined relative to the 1 Kb DNA-ladder. The DNA-band containing the 697 bp long fragment from the digested Gem-MoLV-LTR vector and the DNA-band containing the 4726 bp long fragment from the digested pEGFP-N1 were cut out from the LMP agarose gel and put into two different Eppendorf tubes. Both tubes were briefly centrifuged and incubated at 65 °C for 5 minutes. Afterwards, 3.5  $\mu$ l of the 4726 bp long fragment (Hind III and Eco RI digested pEGFP-N1) were mixed together with 7  $\mu$ l of the 697 bp long fragment (MoLV-LTR promoter from the Hind III and Eco RI digested Gem-MoLV-LTR) in a separate Eppendorf tube, incubated at 65 °C for 5 minutes and centrifuged briefly. The mixture was cooled at 37 °C and then the following ligation mix was added: 2  $\mu$ l 10x ligation buffer, 7.5  $\mu$ l double distilled water and 1  $\mu$ l T4 DNA ligase. The ligation reaction was incubated overnight at 15 °C.

[0065] The newly ligated pEGFP-N1+MoLV-LTR was transformed into One Shot<sup>™</sup> competent *E. coli* (from Invitrogen), according to the manufacturer protocol of Invitrogen. 10  $\mu$ l of the ligation reaction were used for the transformation of competent *E. coli*. 50, 100 and 200  $\mu$ l of the transformation mixture were plated on different kanamycin-containing LB agar plates (final concentration of kanamycin: 50  $\mu$ g/ $\mu$ l) and the plates were then incubated at 37 °C overnight. The next day different bacterial colonies were picked for further analysis of recombinant DNA clones by small-scale plasmid DNA isolation (according to the alkaline lysis method described in Maniatis *et al.*<sup>7</sup>).

[0066] To check the presence of the MoLV-LTR promoter fragment cloned into pEGFP-N1, a F10x incubation buffer for restriction enzyme B, 5  $\mu$ l plasmid DNA from the alkaline lysis method, 22  $\mu$ l double distilled water, 1  $\mu$ l Hind III, 1  $\mu$ l Eco RI were mixed together in an Eppendorf tube and incubated for 3 hours at 37 °C, loaded on a 1% agarose gel and subsequently visualized with 300 nm UV-light and analyzed. The expected fragment of 697 bp length after Hind III and Eco RI digestion was observed, and therefore confirmed the presence of the cloned fragment (see Fig. 3).

[0067] In order to obtain large quantities of the newly created plasmid, a large-scale plasmid DNA isolation has been performed according to the Qia-gen Plasmid Maxi Protocol.

#### Example 3: Electroporation of the murine cell line A20.2J with pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP

[0068] The A20.2J cell line (A20.2J is a BALB/c B cell lymphoma<sup>29-30</sup>, ATCC TIB 208, from American Type Culture Collection) was co-electroporated with the pBluescriptIIKS(+)-EF-1 $\alpha$ -EGFP and pSV2neo (plasmid containing the neo gene which confers resistance to the drug G418). 3x10<sup>7</sup> cells (A20.2J) were required for the electroporation and were plated into fresh medium complete (Iscove's modified Dulbecco's



medium (from Gibco BRL) supplemented with 5 % fetal calf serum (from Gibco BRL), 0.05 mM 2-mercaptoethanol (from Gibco BRL) and 2 mM L-glutamine (from Gibco BRL) one day prior to the poration. Before electroporation, the cells were washed three times in ice-cold  $\text{Ca}^{2+}/\text{Mg}^{2+}$  free PBS (phosphate buffered saline: 8 g NaCl, 0.2 g KCl, 1.44 g  $\text{Na}_2\text{HPO}_4$ , 0.24 g  $\text{KH}_2\text{PO}_4$  in 800 ml double distilled water, adjusted to pH 7.4 with HCl and double distilled water was added to a final volume of 1 liter), counted and  $3 \times 10^7$  cells were resuspended in 800  $\mu\text{l}$  ice-cold PBS. 60  $\mu\text{g}$  pBluescriptIIKS(+)-EF-1 $\alpha$ -GFP and 6  $\mu\text{g}$  pSV2neo were aliquoted into an electroporation cuvette for the co-electroporation of A20.2J. The chilled cells were added to the DNA solution in the electroporation cuvette. The cells and the DNA solution were mixed by pipetting and incubated on ice for 15 minutes. After incubation, the DNA cell suspension was subjected to an electric pulse of 960  $\mu\text{F}$  and 300 V delivered by a BioRad Gene Pulser<sup>TM</sup>. After electroporation, the cells were kept on ice for 15 minutes, followed by a 15 minutes incubation at room temperature. The electroporated A20.2J were then transferred to culture medium (medium complete) and grown for 24 hours before transfectants were selected in medium complete supplemented with G418 (1 mg/ml, from Gibco BRL).

[0069] The electroporated cells were cultured in the selection medium (medium complete supplemented with G418) for 2-4 weeks, analyzed under the fluorescence microscope and then a single-cell-assay was performed in order to obtain a clonal, stable GFP-transfected cell line.

**Example 4: Preparation of cell line PB3cGFP by electroporation of the murine cell line PB-3c with pEGFP-N1+MoLV-LTR**

[0070] The PB-3c cell line (which is a cloned, strictly IL-3 dependent, non tumorigenic mastocyte line isolated from the bone marrow of a DBA/2 mouse<sup>31</sup>) was electroporated with pEGFP-N1+MoLV-LTR.  $3 \times 10^7$  cells (PB-3c) were required for the electroporation and were plated into fresh medium (medium complete supplemented with 2 % IL-3) one day prior to the poration. Before electroporation, the cells were washed three times in ice-cold  $\text{Ca}^{2+}/\text{Mg}^{2+}$  free PBS, counted and  $3 \times 10^7$  cells were resuspended in 800  $\mu\text{l}$  ice-cold PBS. 10  $\mu\text{g}$  pEGFP-N1+MoLV-LTR were aliquoted into an electroporation cuvette for the electroporation of PB-3c. The chilled cells were added to the DNA solution in the electroporation cuvette. The cells and the DNA solution were mixed by pipetting and incubated on ice for 15 minutes. After incubation, the DNA cell suspension was subjected to an electric pulse of 960  $\mu\text{F}$  and 300 V delivered by a BioRad Gene Pulser<sup>TM</sup>. After electroporation, the cells were kept on ice for 15 minutes, followed by a 15 minutes incubation at room temperature. The electroporated PB-3c cells were then transferred to culture

medium (medium complete supplemented with 2 % IL-3) and grown for 48 hours before transfectants were selected in medium complete supplemented with 2 % IL-3 and G418 (1 mg/ml, from Gibco BRL).

[0071] The electroporated cells were cultured in the selection medium for 2-4 weeks, analyzed under the fluorescence microscope and then a single-cell-assay was performed in order to obtain a clonally and stably GFP-transfected cell line named PB3cGFP.

**Example 5: Preparation of the cell line JurkatGFP by electroporation of the human cell line Jurkat with pEGFP-N1+MoLV-LTR**

[0072] The Jurkat cell line (Jurkat is a human acute T cell leukemia from ATCC TIB 152) was electroporated with pEGFP-N1+MoLV-LTR.  $3 \times 10^7$  cells (Jurkat) were required for the electroporation and were plated into fresh RPMI medium (RPMI 1640 medium (from Gibco BRL) supplemented with 5 % fetal calf serum (from Gibco BRL), 0.05 mM 2-mercaptoethanol (from Gibco BRL) and 2 mM L-glutamine (from Gibco BRL)) one day prior to the poration. Before electroporation, the cells were washed three times in ice-cold  $\text{Ca}^{2+}/\text{Mg}^{2+}$  free PBS, counted and  $3 \times 10^7$  cells were resuspended in 800  $\mu\text{l}$  ice-cold PBS. 10  $\mu\text{g}$  pEGFP-N1+MoLV-LTR were aliquoted into an electroporation cuvette for the electroporation of Jurkat. The chilled cells were added to the DNA solution in the electroporation cuvette. The cells and the DNA solution were mixed by pipetting and incubated on ice for 15 minutes. After incubation, the DNA cell suspension was subjected to an electric pulse of 960  $\mu\text{F}$  and 300 V delivered by a BioRad Gene Pulser<sup>TM</sup>. After electroporation, the cells were kept on ice for 15 minutes, followed by a 15 minutes incubation at room temperature. The electroporated Jurkat cells were then transferred to culture medium (RPMI medium) and grown for 48 hours before transfectants were selected in RPMI medium supplemented with G418 (1 mg/ml, from Gibco BRL).

[0073] The electroporated cells were cultured in the selection medium (RPMI medium supplemented with G418) for 2-4 weeks, analyzed under the fluorescence microscope and then a single-cell-assay was performed in order to obtain a clonally and stably GFP-transfected cell line named JurkatGFP.

**Example 6: Comparison of the fluorescent capacities of non-transfected and GFP-transfected cells**

[0074] Three non-related cell lines (A20.2J, PB-3c and Jurkat) were chosen in order to study the fluorescing capacities.

[0075] After stable transfection of the three different cell lines with a GFP-carrying vector, we compared the non-transfected with the GFP-transfected cells of each cell line on behalf of their fluorescing capacities, measured by flow cytometric analysis in a FACScan (Becton

Dickinson, San Jose, USA). The software program Cell Quest (Becton Dickinson, USA) was used to acquire the data. The FACScan was calibrated with Calibrite beads from Becton Dickinson prior to acquisition. The following parameters were measured:

- FSC-Height (forward scatter) as a measure for the cell size,
- SSC-Height (side scatter) as a measure for the internal granularity (density) of the cells,
- green fluorescence (e. g. GFPmut1) was visualised on the FL-1 channel in the FACScan,
- orange fluorescence (e. g. propidium iodide) was visualised on the FL-2 channel in the FACScan.

[0076] For measurements, 10'000 cells were acquired from each FACS-tube containing 250'000 cells.

[0077] There were two surprising findings: The first was the high fluorescence intensity of the GFP-transfected cells compared to the autofluorescence of the non-transfected ones (Fig. 4-6). The second was that the fluorescence-range (visualized on the FL-1 channel in the FACScan) of all tested live GFP-transfected cell lines was narrow (see Fig.): A20GFP (Fig. 4), PB3cGFP (Fig. 5) and JurkatGFP (Fig. 6). This rules out the problem of high and low GFP expressors.

#### Example 7: Apoptosis assay: Induction of apoptosis in A20GFP

[0078] Apoptosis was induced by an 8 hours incubation of A20GFP with the sFasL supernatant (sFasL SN has been produced by FasL-transfected N2a cells, which were provided by A. Fontana, University Hospital, Zurich, Switzerland). As negative control, A20GFP incubated in medium complete has been used.

[0079] After washing A20GFP cells with PBS, two FACS-tubes (from Falcon) were prepared, each containing 250'000 cells. The cells in the first tube (tube 1) were resuspended in 500 µl normal medium complete as negative control, whereas the cells in the second tube (tube 2) were resuspended in 100 µl apoptosis inducing soluble FasLigand supernatant and 400 µl medium complete to a final volume of 500 µl. Both tubes were incubated for 8 hours at 37 °C. After incubation, the cells were washed with PBS, centrifuged at 1400 rpm for 5 minutes, taken up in 500 µl PBS and measured in the FACScan.

[0080] In the control tube (tube 1), the majority of the cells showed a high GFP-associated fluorescence (Fig. 7a and 7b), whereas in the tube containing the sFasL SN treated cells (tube 2) a majority of low fluorescing cells was found after 8 hours incubation (Fig. 8a and 8b). These low fluorescing cells corresponded to the shrunken apoptotic cell population (Fig. 8c and 8e), in contrast to the high fluorescing cells, that are mainly found in the bigger and less granular living cell population (Fig. 8c and 8d).

[0081] To further show and confirm, that the low fluorescing cell population corresponds to the apoptotic shrunken cell population, a propidium iodide staining for intracellular DNA has been performed with the remaining cells of the previously measured tubes (tube 1 and 2). The cells were first washed with PBS, then incubated in 70 % ethanol at -20 °C for 2 hours, washed again with PBS and stained with 500 µl PI-solution (50 µg PI in 1 ml PBS) for 30 minutes at 37°C. Both tubes (tube 1 and 2), containing PI stained cells, were immediately measured in the FACScan.

[0082] The DNA profile of the apoptosis inducing sFasL SN treated cells (tube 2) was considerably changed after 8 hours incubation (figures 10a and 10b). The majority of the DNA has been fragmented (visualized as sub-G1 peak on the FL-2 channel in a FACScan, figure 10b), in comparison to the control (tube 1, see figures 9a and 9b), confirming the result of massive apoptosis induced by sFasL SN.

[0083] Furthermore, the cell population with fragmented DNA corresponded to the group of the shrunken apoptotic and low fluorescing cells.

[0084] In conclusion, apoptotic cells shrink, they have mainly (but not completely) lost the GFP-associated fluorescence and have a fragmented DNA (which is a specific characteristic of apoptosis). Moreover, the amount of apoptotic cells seen with the PI method was comparable to the amount detected with the GFP method (data not shown).

#### Example 8: Toxicity assay: Induction of necrosis in A20GFP

[0085] A20GFP cells were first washed with PBS and then 250000 cells were incubated in a FACS-tubes with 100 µl anti-A20 polyclonal antibody for 60 min at 4°C. After incubation, the cells were washed with PBS, resuspended in 50 µl rabbit complement (Readsystem) and incubated in a 37 °C waterbath for 1 hour. The cells were washed again with PBS and resuspended in 500 µl PBS and immediately measured in a FACScan. As controls, untreated (without complement) A20GFP and A20 cells were used.

[0086] Complement binding induces necrosis in target cells. This assay was thus performed in order to evaluate the fluorescing capacity of necrotic cells. The findings were striking, as necrotic cells completely lost their GFP-associated fluorescence capacity within an hour. Thus complement-treated A20GFP cells can hardly be distinguished from non-transfected live A20 cells (Fig. 11). The necrotic cells can be distinguished from apoptotic cells, the latter still express low fluorescing capacity (e. g. 8 hours after sFasL SN treatment, as shown in the example described before). This makes the GFP-assay a useful tool for the discrimination between necrotic, apoptotic and living GFP-transfected cells (Fig. 11).

**Example 9: Inhibition of de novo RNA synthesis by actinomycin D (ActD), protein synthesis by cycloheximide (CHX) and the potential role of interfering with the GFP apoptosis assay in A20GFP**

[0087] To find out whether the fluorescence of A20GFP and the GFP apoptosis assay as described in example 7 is dependent on the de novo transcription and translation of proteins, Act D (pharmacy of the University Hospital, Basel), a RNA synthesis inhibitor, and Cycloheximide (Sigma), a protein synthesis inhibitor, were tested in our experimental protocol. After washing A20GFP cells with PBS, 250'000 cells were aliquoted in FACS-tubes and incubated with:

- a) Tube 1 (control): 500 µl medium complete (MC),
- b) Tube 2: 100 µl sFasL SN and 400 µl MC,
- c) Tube 3: 500 µl MC and 5 µg/ml ActD,
- d) Tube 4: 400 µl MC, 100 µl sFasL SN, 5 µg/ml ActD,
- e) Tube 5: 500 µl MC and 10 µg/ml CHX,
- f) Tube 6: 400 µl MC, 100 µl sFasL SN and 10 µg/ml CHX;
- g) Tube 7: 500 µl MC, 5 µg/ml Act D and 10 µg/ml CHX
- h) Tube 8: 400 µl MC, 100 µl sFasL SN, 5 µg/ml Act D and 10 µg/ml CHX.

[0088] Tubes 1-8 (a-h) were incubated for 10 hours at 37 °C. The fluorescing status of the cells was assayed at 10 hours post treatment.

[0089] As seen in figures 12a and 12b, there is no difference between the fluorescence intensity of untreated A20GFP (Fig. 12a) compared to A20GFP treated with ActD and/or CHX (Fig. 12b). sFasL SN treatment of A20GFP induces apoptosis to the same extent, whether the cells were ActD- and/or CHX-treated or not. In conclusion, ActD and CHX, either alone or together, do not substantially interfere with the fluorescing capacity of A20GFP nor do they inhibit sFasL SN induced apoptosis after 10 hours of treatment. Therefore, A20GFP, in order to fluorescence, do not require de novo transcription and translation during the 10 hours.

**Example 10: Apoptosis assay in PB3cGFP by Interleukin-3 deprivation**

[0090] PB3cGFP is an interleukin-3 (IL-3) dependent cell line. The addition of IL-3 to the culture medium (medium complete) prevents the PB3cGFP cells from undergoing apoptosis (IL-3 as an anti-apoptotic compound). In contrast, upon IL-3 deprivation a strong apoptotic response is seen within 12 hours.

[0091] For this experiment, 250'000 PBS-washed PB3cGFP cells were aliquoted in two different FACS-tubes.

Tube A: contained the cells and MC supplemented

with IL-3.

Tube B: contained the cells and MC without IL-3.

[0092] Both tubes were incubated for 12 hours at 37 °C. The GFP-associated fluorescence of the PB3cGFP cells was measured after 12 hours of IL-3 deprivation.

[0093] As expected, and seen with A20GFP, very similar results were obtained with PB3cGFP. PB3cGFP, cultured in MC supplemented with IL-3 (tube A), showed a high GFP-associated fluorescence, whereas IL-3 deprived PB3cGFP cells (tube B) underwent apoptosis and demonstrated a low GFP-associated fluorescence intensity, making them clearly distinguishable from live PB3cGFP and live PB3c cells (Fig. 13). These findings have been confirmed by the PI staining method, as it has been described with the A20GFP cells (data not shown).

**Example 11: Apoptosis assay: Induction of apoptosis in JurkatGFP**

[0094] This apoptosis assay has also been performed with sFasL SN, a known cell death inducer in Jurkat cells. 250'000 PBS-washed JurkatGFP cells were aliquoted in two different FACS-tubes.

Tube 1: (control): cells were incubated with 500 µl RPMI medium,

Tube 2: cells were incubated with 100 µl sFasL SN and 400 µl RPMI medium

[0095] Both tubes were incubated for 18 hours at 37 °C. In this experiment cells were immediately measured after incubation, without prior washing with PBS.

[0096] As expected, and seen with A20GFP and PB3cGFP cells, very similar results were obtained.

[0097] As shown in Fig. 14, sFasL SN induced apoptosis. GFP-associated fluorescence was high in live JurkatGFP, whereas the apoptotic JurkatGFP cells had a marked reduced GFP-associated fluorescence, but were still distinguishable from non-transfected live Jurkat cells. These findings have been confirmed by the PI staining method.

[0098] In relation to the previous experiments, the present results show that it is not necessary to wash the cells with PBS before measuring. This may be particularly useful for large screening assays, where individual tubes cannot be separately washed.

#### Abbreviations

[0099] The following abbreviations are used throughout the description:

ActD	Actinomycin D
bp	base pairs
CHX	Cycloheximide
DIG	Digoxigenin

ds DNA	double-stranded DNA	
EDTA	Ethylenediaminetetraacetic acid	
EF-1a	elongation factor 1a	
FACS	Fluorescence Activated Cell Sorter	
FITC	Fluorescein isothiocyanate	5
FL-1	Fluorescence channel 1 (detects fluorescence emission in the range of 530 (+/-25) nm	
FSC	Forward Scatter (cell size)	
GFP	Green Fluorescent Protein	10
IL-3	Interleukin-3	
LMP	low melting point	
MC	medium complete	
PBS	phosphate buffered saline	
PI	propidium iodide	15
sFasL	soluble Fas Ligand	
SN	supernatant	
SSC	Side Scatter (granularity of cells)	
ss DNA	single-stranded DNA	
TAE	Tris(hydroxymethyl)aminoethane	20
TNF	Tumor Necrosis Factor	
TUNEL	TdT-mediated end labelling of DNA	

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[0100]

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## Claims

1. A method for the determination of apoptotic and/or necrotic conditions of living test cells comprising monitoring the presence of a marker protein in said cells.
2. A method according to claim 1, wherein the apoptotic and/or necrotic condition is monitored in the presence of a non-, pro-, or anti-apoptotically or necrotically active compound and/or a physical stimulus.
3. A method according to claim 1, wherein the marker protein is produced in the test cells after transfection of said cells with a DNA coding for and expressing the marker protein.
4. A method according to claim 1, wherein the marker protein is the green fluorescence protein (GFP) or a fluorescent mutant thereof.
5. A method according to claim 1, wherein the presence of GFP or the fluorescent mutant thereof is monitored by means of a flow cytometer or a plate reader measuring the fluorescence intensity.
6. A method according to claim 1, wherein two groups

- of test cells are used, each of a defined number of cells, which were transfected with a DNA vector coding for a fluorescent marker protein, such as the GFP or a fluorescent mutant thereof, incubating one group together with the test compound in a culture medium, stimulating the cells of both groups with an excitation beam, determining the fluorescing intensities of the cells of each group by means of a flow cytometer, and comparing the obtained fluorescing intensity of the cells of the two groups.
7. A method according to claim 6, wherein the DNA vector pBluescriptIIKS(+)+EF-1 $\alpha$ +EGFP or pEGFP-N1+MoLV-LTR are used.
  8. A new vector for the stable transfection of test cells which is sufficiently stable within the test cell and able to highly express a marker protein.
  9. A vector according to claim 8, comprising a gene coding for the marker protein which is operably linked to one or more strong promoters, such as the hEF1- $\alpha$  promoter, the MoLV-LTR promoter or a combination of the CMV and the MoLV-LTR promoter.
  10. A vector according to claim 8 wherein the gene codes for the GFP or a fluorescent mutant thereof.
  11. A vector according to claim 8 which is able to express the marker protein GFPmut1.
  12. A vector according to claim 8 which is pBluescriptIIKS(+)+EF-1 $\alpha$ +EGFP or pEGFP-N1+MoLV-LTR.
  13. A live cell line transfected with a vector according to anyone of claims 8 to 12.
  14. A living cell line transfected with a vector according to claim 12 which is A20GFP, PB3cGFP or Jurkat-GFP.
  15. A method to assay the non-, pro- or anti-apoptotic or necrotic activity of a test compound and/or of a physical stimulus in living test cells comprising monitoring the presence of a marker protein in said cells.
  16. A method according to claim 15, comprising transfecting a group of said cells with a vector coding for and expressing a marker protein, treating the transfected cells in a suitable culture medium with the test compound, monitoring the presence of the expressed marker protein in said group of cells and comparing the results with the results observed with a parallel group of the same test cells which was not treated with the test compound.
  17. A method according to claim 15, wherein the test compound comprises a multiplicity of compounds, e. g. as obtained from combinatorical chemistry methods.
  18. A method according to claim 15, wherein normal, infected or cancer cells are used.
  19. A method according to claim 15, wherein as test cell transfected cell lines A20GFP, PB3cGFP or JurkatGFP are used.
  20. A method according to claim 15, wherein the test cells are transfected with a vector according to any one of claims 8 to 12.
  21. A method according to claim 15, wherein the test cells are transfected with the vector pBluescriptIIKS(+)+EF-1 $\alpha$ +EGFP or pEGFP-N1+MoLV-LTR.
  22. A method according to claim 15, wherein monitoring is performed with the aid of a flow cytometer, e. g. the FACScan<sup>TM</sup>.
  23. A method according to claim 15, wherein monitoring is carried out by measuring the parameters FSC-Height, SSC-Height and the fluorescence of the marker protein and comparing the results after dot plot and/or histogram visualisation.

Figure 1:

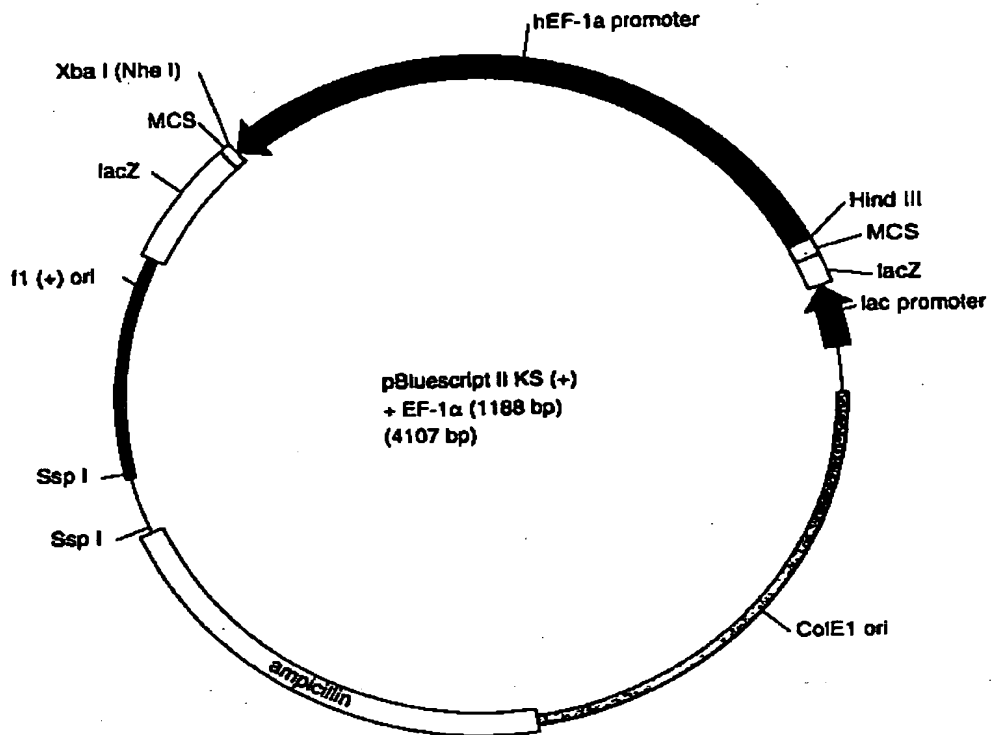


Figure 2:

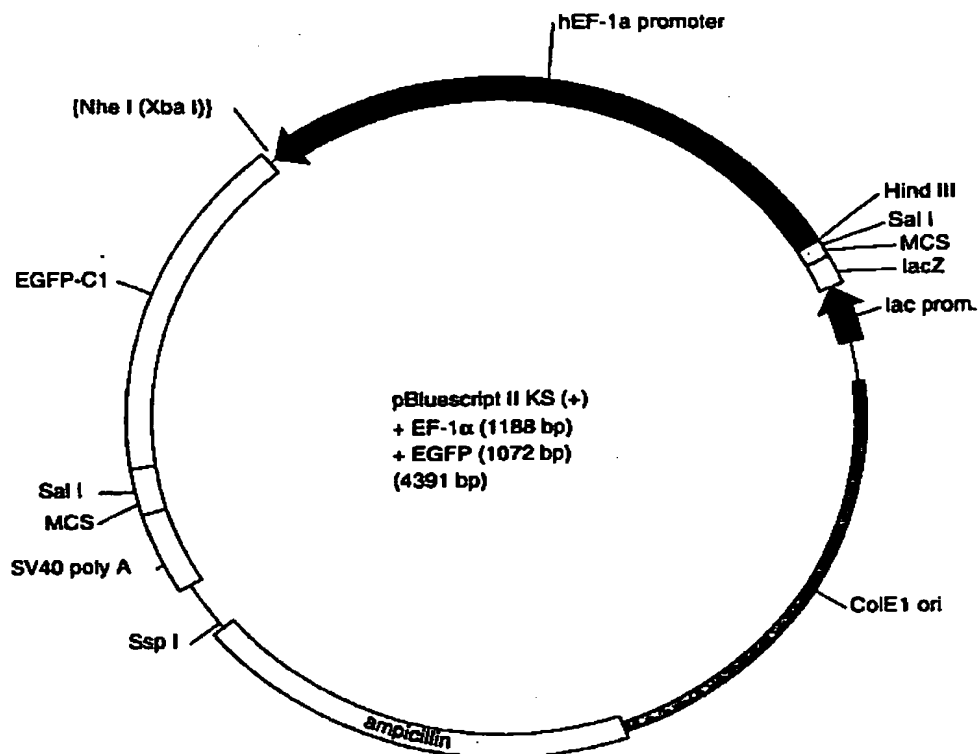


Figure 3:

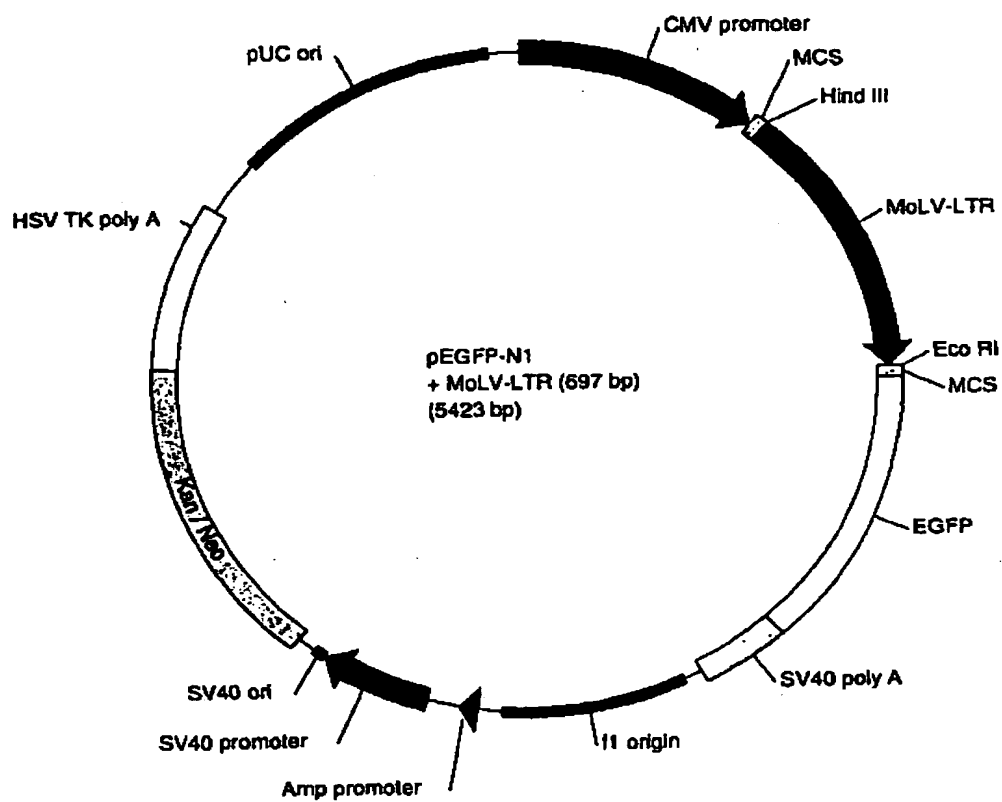




Figure: 4

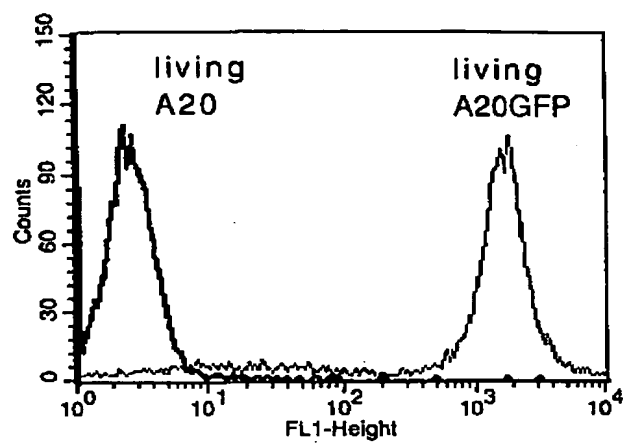


Figure: 5

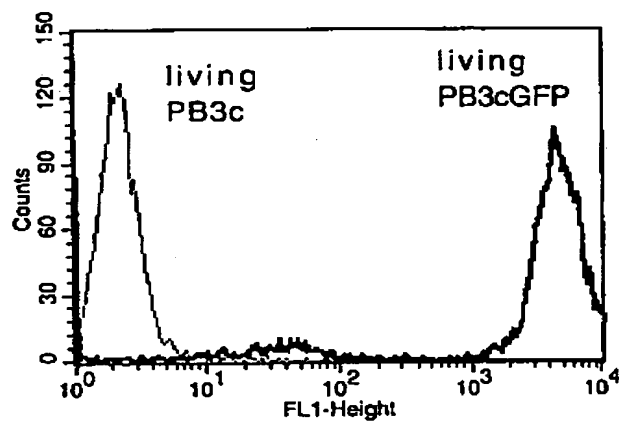


Figure: 6

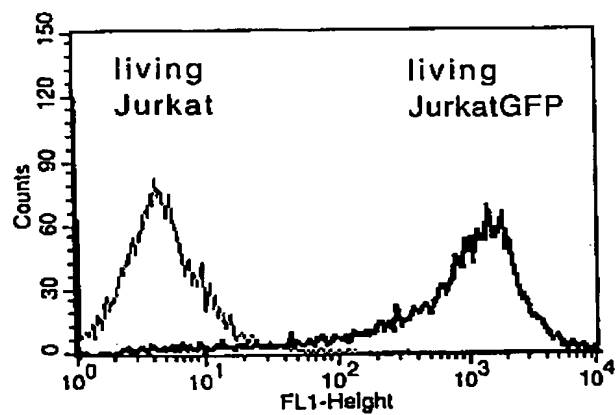


Figure: 7a

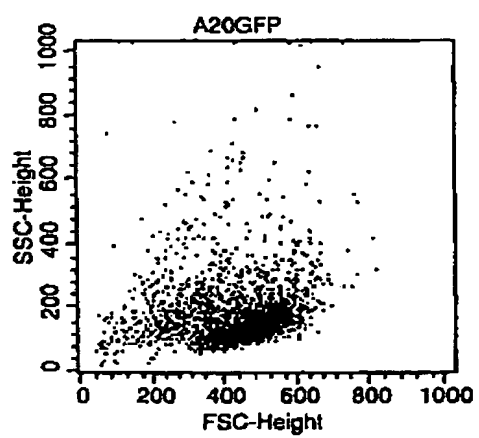


Figure: 7b

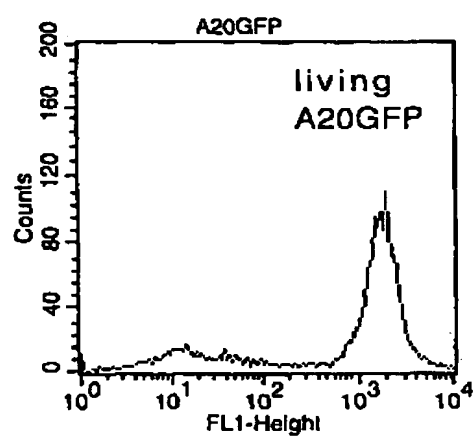


Figure: 8a

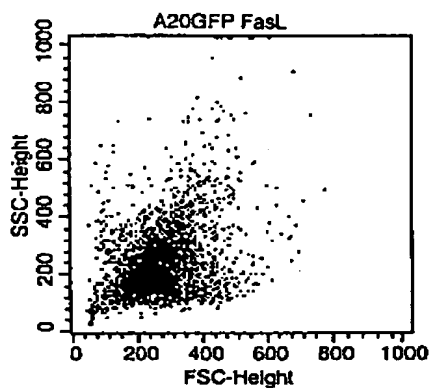


Figure: 8b

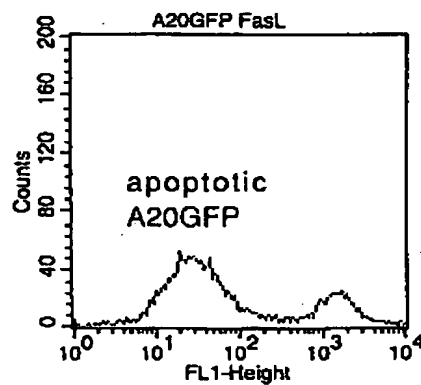


Figure: 8c

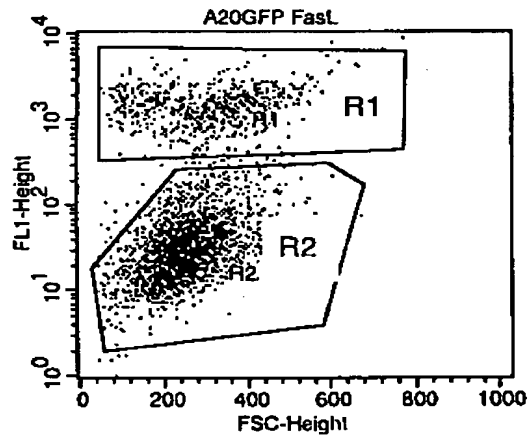


Figure: 8d

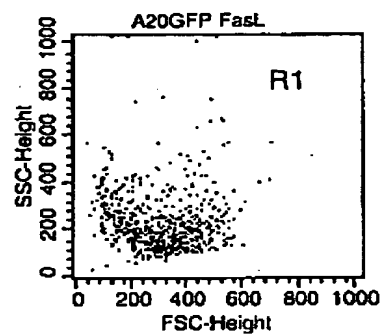


Figure: 8e

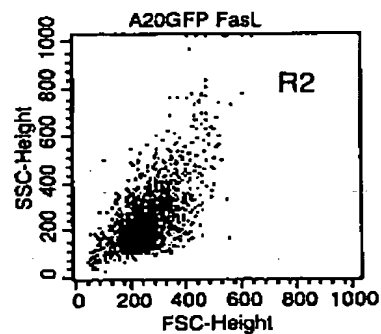


Figure: 9a

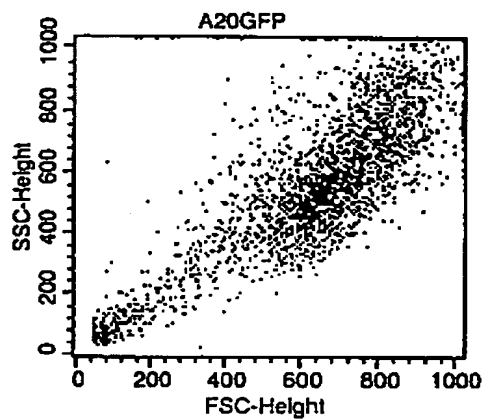


Figure: 9b

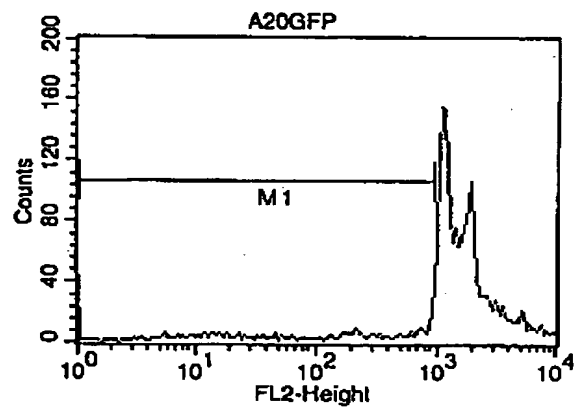


Figure: 10a

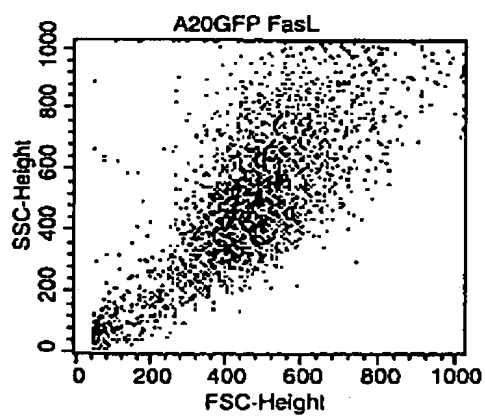


Figure: 10b

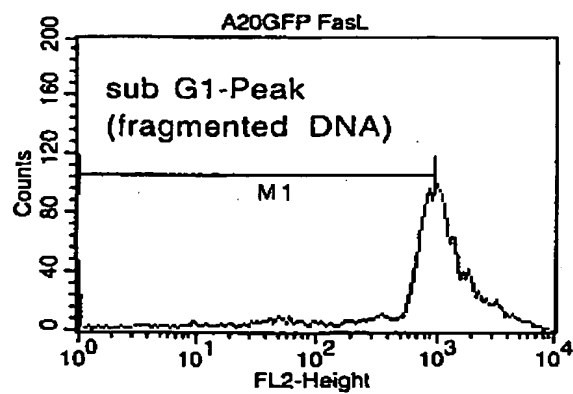


Figure: 11

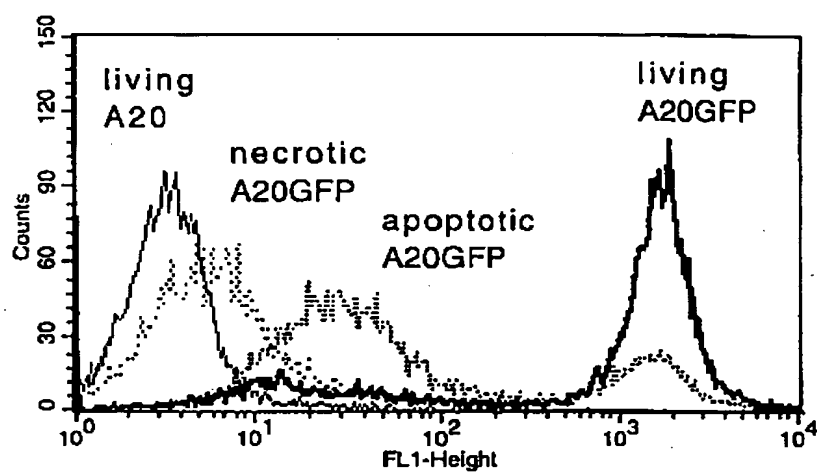


Figure: 12a

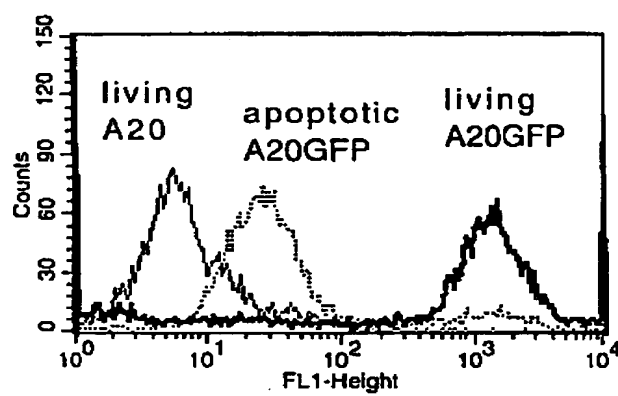


Figure: 12b

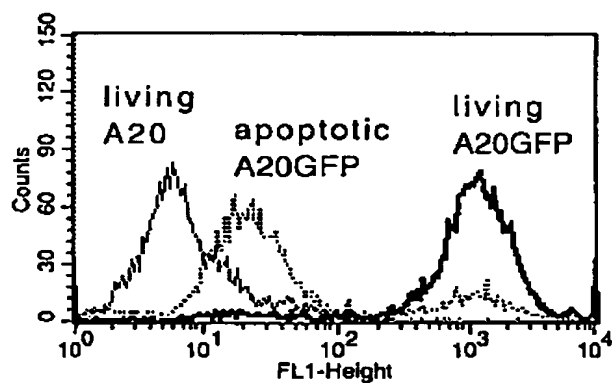


Figure: 13

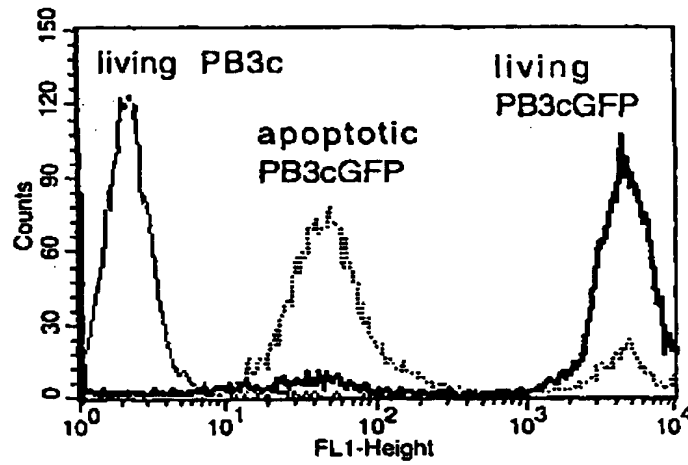
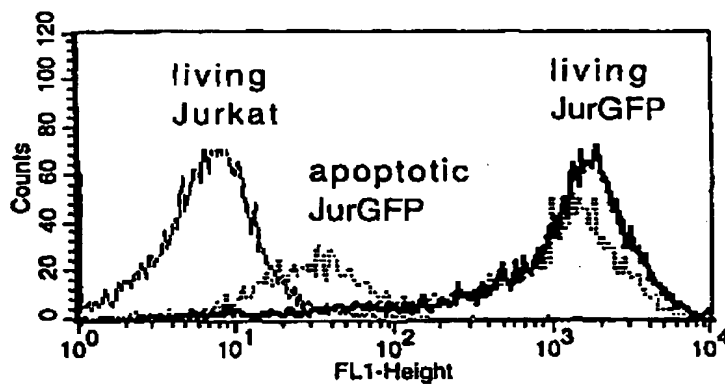


Figure: 14





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# EUROPEAN SEARCH REPORT

Application Number  
EP 98 10 0369

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. CL. 8)
X	WO 97 14812 A (CHIRON CORP) 24 April 1997  * claims 1,51,55-57 *	1-6,8, 10,11, 13-20, 22,23	G01N33/50 C12N15/58 C12Q1/68
X	WO 97 11094 A (NOVONORDISK AS ;THASTRUP OLE (DK); TULLIN SOEREN (DK); POULSEN LAR) 27 March 1997  * page 7, line 9 - line 13 *	1,6,8, 10,11, 13-20, 22,23	
X,D	LAMM, G.M. ET AL.: "A rapid, quantitative and inexpensive method for detecting apoptosis by flow cytometry in transiently transfected cells." NAR, vol. 25, no. 23, 1997, pages 4855-4857, XP002068178 * the whole document *	1,6,8, 10,11, 13-20, 22,23	
X	Y. LI AND M.S. HORWITZ: "Use of a green fluorescent protein in studies of apoptosis of transfected cells." BIOTECHNIQUES, vol. 23, no. 6, 1997, pages 1026-1029, XP002068179 * the whole document *	1-6,8, 10,11, 13-20, 22,23	TECHNICAL FIELDS SEARCHED (Int. CL. 8)  G01N C07K C12N C12Q
X	EP 0 806 668 A (BECTON DICKINSON CO) 12 November 1997 * claim 1 *	1	
-/-			
The present search report has been drawn up for all claims			
Place of search <b>THE HAGUE</b>		Date of completion of the search <b>16 June 1998</b>	Examiner <b>Hoekstra, S</b>
<p><b>CATEGORY OF CITED DOCUMENTS</b></p> <p>X : particularly relevant if taken alone  Y : particularly relevant if combined with another document of the same category  A : technological background  O : non-written disclosure  P : intermediate document</p> <p>T : theory or principle underlying the invention  E : earlier patent document, but published on, or after the filing date  D : document cited in the application  L : document cited for other reasons  &amp; : member of the same patent family, corresponding document</p>			

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# EUROPEAN SEARCH REPORT

Application Number  
EP 98 10 0369

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
A	CORMACK B P ET AL: "FACS-optimized mutants of the green fluorescent protein (GFP)" GENE, vol. 173, no. 1, 1996, page 33-38 XP004042850 * the whole document *	1-23	
A	WO 95 27903 A (INSTITUUT BIOPRIME ;REUTELINGSPERGER CHRISTIAAN PE (NL)) 19 October 1995 -----	1-23	
			TECHNICAL FIELDS SEARCHED (Int.Cl.6)
The present search report has been drawn up for all claims			
Place of search <b>THE HAGUE</b>		Date of completion of the search <b>16 June 1998</b>	Examiner <b>Hoekstra, S</b>
<b>CATEGORY OF CITED DOCUMENTS</b> X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	

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